

Calmodulin is necessary for vegetative growth, ultraviolet survival, and sexual development in the model filamentous fungus *Neurospora crassa*

Calcium (Ca^{2+}) is an important player of intracellular signaling system in living organisms including fungi. In the model filamentous fungus *Neurospora crassa*, calmodulin (CaM) is a high affinity Ca^{2+} -binding protein involved in the Ca^{2+} signaling process to regulate numerous cell functions. The *N. crassa* CaM is highly similar to its human homologue, however, it is less acidic than its vertebrate counterpart with amino acid composition is typical to that of plant homologues. CaM is an essential gene in *N. crassa*, deleting the CaM gene or its direct knockout would be lethal. Therefore, in a previous study, we used CaM antagonists trifluoperazine (TFP) and chlorpromazine (CPZ) to study the functions of CaM in *N. crassa*.

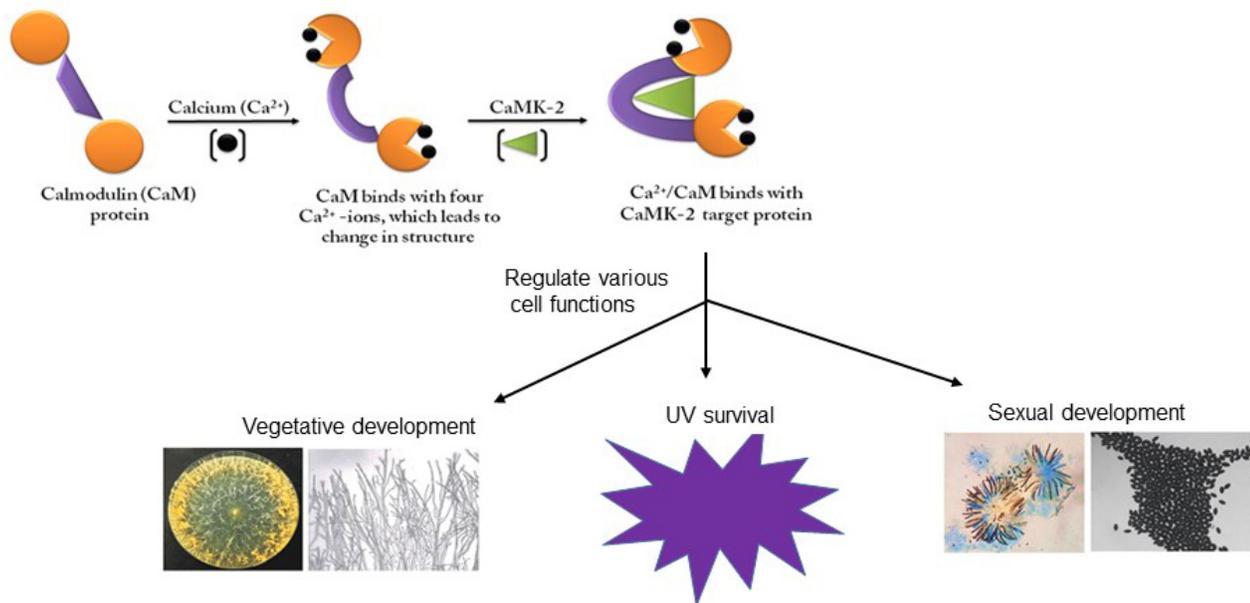


Fig. 1. Overview of CaM and Ca^{2+} /CaMK-2 function in *N. crassa*. CaM is a dynamic dumbbell-shaped Ca^{2+} -binding protein, which responds to a wide range of Ca^{2+} concentrations ($\sim 10^{-12}$ M - 10^{-6} M), binding of Ca^{2+} (black dots) changes the CaM protein conformation and charge. CaM binds to different target enzymes and proteins to amplify the Ca^{2+} -signaling cascade, including a Ser/Thr kinase Ca^{2+} /CaMK-2 (green triangle) to perform various cell functions including vegetative growth, UV survival, and sexual development.

In the Ca^{2+} signaling process, CaM interacts with target proteins including the conserved Ca^{2+} /CaM-dependent Ser/Thr protein kinase 2 (Ca^{2+} /CaMK-2). Like other Ca^{2+} /CaMK-2 proteins,

its *N. crassa* homologue also contains a conserved N-terminal catalytic domain and a C-terminal regulatory domain containing overlapping autoinhibitory and Ca²⁺/CaM binding. Binding of Ca²⁺/CaM displaces the autoinhibitory segment and activates the kinase, and then the kinase lock itself into activated states by autophosphorylation on a conserved threonine residue in the autoinhibitory segment that increases its affinity for CaM to about 1000-fold. We generated *N. crassa* strains duplicated for the CaM gene. In *N. crassa*, duplicated DNA sequences are the target of repeat-induced point mutation (RIP) that causes multiple G:C to A:T mutations and methylation of remaining cytosine residues within both copies of the duplicated DNA sequences after fertilization but before karyogamy during the sexual cycle. Because, *N. crassa* produces thousands of ascospores in the sexual cycle, it is possible to screen a large number of progeny for various RIP-induced mutations in the CaM. However, because CaM is essential, we expected to isolate only the CaM^{RIP} strains containing viable mutations in the CaM gene. We isolated a RIP-induced CaM mutant, designated as *cmd*^{RIP}(26), that vegetative phenotypes such as slow growth rate, reduced aerial hyphae development, and reduced accumulation of carotenoid that is responsible for the characteristic orange pigmentation in *N. crassa*, than the wild type strain. Moreover, the viability of *cmd*^{RIP}(26) was severely reduced on exposures to ultraviolet (UV) irradiations. Furthermore, we found that the product of the CaM during meiosis was essential for full fertility in *N. crassa*. In addition, in the *N. crassa* Ca²⁺/CaMK-2 protein, we introduced S247A and T267A mutations in the candidate phosphorylation sites in the catalytic domain, and L309D mutation in the predicted CaM binding domain, and studied the phenotypes of these mutant strains. We found that the crosses homozygous for the *camk-2*^{S247A} and *camk-2*^{T267A} showed an intermediate phenotype (produced few hundreds of ascospores), indicating that phosphorylation of serine 247 and threonine 267 residues play a role in full fertility in *N. crassa*. In addition, *cmd* gene was upregulated in the strains lacking the Ca²⁺/CaMK-2, which might suggest that CaM might compensate the lack of Ca²⁺/CaMK-2 in *N. crassa*. Thus, in this study, CaM has been found necessary for normal vegetative and sexual developments, and UV survival. In addition, serine 247 and threonine 267 residues were found important for the Ca²⁺/CaMK-2 functions in *N. crassa*.

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